Baseline Plasma Total Homocysteine and Adenoma Recurrence: Results from a Double Blind Randomized Clinical Trial of Aspirin and Folate Supplementation

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Abstract

Background: Elevated plasma total homocysteine (tHcy) is an accepted marker of functional folate deficiency but may have independent effects on colorectal neoplasia risk. It is uncertain whether plasma tHcy is associated with risk at the low levels common in a folate-fortified population.

Methods: Study subjects, about half of whom were recruited after fortification of grain products with folic acid in the United States and Canada, consisted of 871 individuals with a recent history of one or more colorectal adenomas who were randomized to receive either a 1 mg/day folic acid supplement or a placebo within one of three randomly assigned aspirin treatment groups (placebo, 81, or 325 mg/day). Nonfasting plasma tHcy was determined by a gas chromatograph mass chromatography method. We estimated adjusted risk ratios and 95% confidence intervals (95% CI) for one or more adenoma recurrences for each quartile of baseline plasma tHcy using generalized linear regression with an overdispersed Poisson approximation to the binomial.

Results: The Q4/Q1 adjusted risk ratio for any adenoma was 0.98 (95% CI, 0.70-1.38; *P* trend = 0.17) in the placebo group, and 0.81 (95% CI, 0.58-1.12; *P*-trend = 0.17) in the folic acid group. Results were similar for adenomas with advanced features. There was no modification by sex, aspirin treatment group or *MTHFR* 677C>T genotype.

Conclusions: Plasma tHcy is not an independent marker for an increase in colorectal adenoma recurrence risk in postfortification populations in which plasma tHcy levels are in the lower range of values.

Impact: Controlling plasma tHcy levels is unlikely to favorably modify adenoma recurrence risk in folate-fortified populations. *Cancer Epidemiol Biomarkers Prev;* 19(10); 2541–8. ©2010 AACR.

Introduction

Plasma total homocysteine (tHcy) is a sulfur-containing amino acid and a product of *S*-adenosylmethionine– dependent methylation reactions. Plasma tHcy is a marker of overall folate-dependent one-carbon meta-

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bolism because it reflects the status of several of the B vitamins and methyl donors involved. Although plasma folate is the main determinant of plasma tHcy, the B vitamins B_{12} , B_6 , and B_2 are also significant determinants (1-4) as is the *MTHFR* genotype (5-8).

Interest in plasma tHcy initially stemmed from its association with cardiovascular disease risk (9). Much less is known about the association between plasma tHcy and cancer. Plasma tHcy levels are associated with potential cancer risk factors, such as cigarette smoking and body mass index, only in part because of their association with plasma folate (1-4, 10). An effect of plasma tHcy on carcinogenesis independent of B-vitamin status is also possible, given that it is in equilibrium with *S*-adenosylhomocysteine (SAH), a potent inhibitor of intracellular methylation reactions (11-13), and that elevated plasma tHcy may be associated with increased DNA damage even after controlling for plasma folate (14, 15).

An association between increased plasma tHcy and risk of colorectal neoplasia is suggested by four studies, all of which reported similarly increased risks for colorectal cancer or adenomas in subjects with the highest plasma tHcy (12, 16-18). However, only one study (12) controlled for plasma folate and other plasma tHcy determinants,

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Note: Author contributions: A.J. Levine and J.A. Baron conceived of the study, and participated in its design and in drafting the manuscript. M.V. Grau and L.A. Mott structured and carried out the statistical analyses. P.M. Ueland supervised the laboratory analyses of plasma tHcy and B-vitamins and assisted in drafting the manuscript. All authors read and approved the final manuscript.

The folic acid and placebo tablets used in this clinical trial were provided by Wyeth.

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and all studies involved subjects with relatively low folate intake. Thus, it is not clear whether the association between plasma tHcy and colorectal neoplasia is independent of folate availability or whether the association is still present when plasma tHcy is in the lower range. Plasma tHcy was not associated with colorectal cancer risk in three more recent studies (19-21).

In the current study we assessed the association between plasma tHcy and subsequent colorectal adenoma recurrence risk in subjects who were randomized to receive either a 1 mg/day folic acid supplement or a placebo for up to three years. In our study population, subjects were randomized and followed during a period of increasing folic acid intake in the source population, during which time the mean plasma tHcy levels in the study population decreased. We sought to assess how any associations with baseline plasma tHcy were modified by folic acid treatment assignment. We hypothesized that if plasma tHcy was a risk marker for adenoma recurrence it would be evident only in the placebo group, which maintained higher mean plasma tHcy levels during follow-up than those in the folic acid group in which plasma tHcy remained uniformly low.

Materials and Methods

Design

These data were collected as part of a randomized, double-blind, placebo-controlled trial of the efficacy of oral aspirin (ASA), folic acid, or both to prevent colorectal adenomas as described (22, 23). Briefly, this was a three-by-two factorial design in which subjects were randomized to receive 81 mg/day ASA, 325 mg/day ASA, or placebo. Within each ASA/placebo group, subjects were additionally randomized to receive 1 mg supplemental folic acid/day or a placebo. The study initially focused only on aspirin; 100 subjects who were randomized only to aspirin were not included in this analysis.

Recruitment, randomization, treatment, and follow-up

Details of subject eligibility, recruitment, randomization, treatment, and follow-up, and study outcomes have been described (22, 23). Briefly, subjects were recruited from July 1994 until March 1998 from nine clinical centers. Eligible subjects were between 21 and 80 years of age, in good health, and had received a recommendation for a 3-year follow-up colonoscopy by their regular medical practitioner. Each eligible subject met at least one of the following three criteria: (a) one or more histologically confirmed colorectal adenomas removed within three months of their recruitment; (b) one or more histologically confirmed adenomas removed within 16 months of their enrollment as well as a history of two or more confirmed adenomas; or (*c*) an adenoma >1 cm in diameter. Individuals were ineligible if they had a history compatible with a familial colorectal cancer syndrome, invasive

colorectal cancer, any malabsorption syndrome, a medical condition that could be worsened by use of aspirin or folic acid, or any medical condition commonly treated with aspirin, nonaspirin nonsteroidal anti-inflammatory drugs, or folate. Before entering the trial all subjects were required to have had a complete colonoscopy with removal of all polyps within three months of their entry into the trial. Each subject underwent a three-month run-in period on 325 mg ASA prior to randomization into ASA and folic acid treatment groups. Only subjects with \geq 80% compliance and no other contraindications were randomized. By protocol, all subjects had an anticipated followup complete surveillance colonoscopy 34 to 40 months after the qualifying examination.

In the initial study design, subjects in each treatment group were followed for three years and adenoma recurrence was determined at the end of that three-year follow-up interval (the first follow-up). Due to concern that a longer follow-up interval might be necessary to observe the effect of folic acid supplementation, subjects were invited to remain on the study supplements until the next surveillance examination (typically three or five years later). The current analysis reflects plasma tHcy measurements made at study enrollment and adenomas found during the first follow-up period, i.e., through 36 to 40 months after the baseline colonoscopy. To assess the change in plasma tHcy levels over the follow-up interval, a second measurement was taken at the beginning of the second follow-up.

Study outcomes

The primary study outcome was the proportion of patients in whom one or more colorectal adenomas were detected in the period starting one year after randomization to the end of the year 3 surveillance follow-up examination. If a year-3 colonoscopy was not done, we used the last examination at least one year after randomization. Adenomas were classified as neoplastic (adenomatous) or nonneoplastic by the study pathologist, who also assessed the degree of dysplasia and the extent of villous component in each adenoma. We defined advanced lesions as invasive carcinoma or adenomas with at least 25% villous component, high-grade dysplasia, or an estimated size of ≥ 1 cm. Patients were considered to have "multiple adenomas" when there were a total of ≥ 3 follow-up adenomas by the end of the year 3 exam.

Plasma total Hcy, folate, and vitamin B_{2r} B_{6r} and B_{12} determination

Plasma tHcy and other nutrients were determined in nonfasting blood samples taken at baseline and approximately three years later. Blood samples were collected into 7-mL Vacutainer brand tubes containing EDTA. After collection, specimens were immediately put on ice and then centrifuged at $1,100 \times g$ for 10 minutes. Whole blood, plasma, and buffy coat fractions were stored at -20° C and then transferred to Dartmouth Medical School where they were stored at -80° C until analysis. Plasma

tHcy was determined by a gas chromatograph mass chromatography method as described (24). Plasma total folate and vitamin B_{12} were determined by microbiological assay using published methods (25, 26). Vitamins B_6 (as PLP, the main active form of vitamin B_6) and B_2 were determined in plasma by liquid chromatography-tandem mass spectrometry (27). All B-vitamin assays were conducted at the laboratory of Bevital AS, Bergen Norway.

Questionnaires

Subjects completed a questionnaire at baseline that queried smoking history, height, weight, and personal medical history. Dietary variables as well as use of multivitamins, coffee and tea consumption, and alcohol use were derived from the Block food frequency questionnaire (28) and reflected average intake over the prior 12 months. Daily nutrient intakes were estimated using a nutrient data base and subject estimates of the number of servings per day multiplied by the estimated serving size for each food.

MTHFR genotype

The *MTHFR* polymorphisms, 677C>T and 1298A>C, were genotyped with the 5'nuclease TaqMan allelic discrimination assay using the ABI7900 (Applied Biosystems). PCR primers and dual-labeled allele discrimination probes were designed using the Primer Express software package (PE Biosystems) as described by Gibson et al. (29). Each 384-well assay contained internal quality controls for homozygous wild-type, heterozygous, and homozygous variant alleles for the respective polymorphisms along with no-template controls. Genotype calls were determined by SDS 2.1 analysis software.

Statistical analysis

Statistical significance was defined as a two-sided P value ≤ 0.05 . To compare those with an adenoma recurrence with those without a recurrence, we used *t*-tests or Wilcoxon rank-sum tests for continuous variables and χ^2 tests for categorical variables. The main exposure variable was quartile of baseline plasma tHcy. Baseline plasma tHcy quartiles were determined with reference to the whole study population.

In the analysis of the change in plasma tHcy over calendar time we calculated means and SD by recruitment year and estimated a *P* for trend using orthogonal linear contrasts and Wald tests. For the analysis of the association between plasma tHcy quartile and number of polyps at the baseline examination we used a proportional odds model, where the outcome was the number of baseline adenomas (1, 2, 3, 4 or \geq 5; ref. 30). The output of this analysis can be interpreted as the odds ratio (OR) for the higher versus lower adenoma number, whatever the high/low cut point. We controlled for *a priori* potential confounders by adding to the model variables that have been associated with both plasma tHcy and adenoma recurrence in the literature or in the current trial population: age, cigarette smoking (never, former current), alcohol use (continuous, as drinks/day), body mass index (BMI; continuous), plasma total folate, plasma B_2 , plasma B_6 (as PLP), and plasma B_{12} (all as continuous values). Race/ethnic group was not a confounder in this population, which was over 85% non-Hispanic white. We did not measure serum creatinine. We used the baseline measures for all of the potential confounders in this analysis.

In the analysis of the effect of plasma tHcy on adenoma recurrence we estimated risk ratios (RR) and 95% confidence intervals (95% CI) for one or more adenomas after randomization with a generalized linear regression analysis using an overdispersed Poisson approximation to the binomial. We obtained *P* values for trend using orthogonal linear contrasts. In the analysis we controlled for age, sex, study center, alcohol, BMI, smoking, plasma levels of plasma total folate, B₂, B₆, and B₁₂, and aspirin treatment group, as defined above. Separate models were used to assess the association between plasma tHcy and recurrence risk within groups defined by folate and aspirin treatment assignment, sex, and the MTHFR 677C>T genotype. We used multiplicative interaction terms and Wald tests to test for interaction. STATA version 9.2 was used for all analyses.

Results

There were 1,021 subjects randomized to folic acid or placebo. A total of 871 subjects (85.0%) had baseline and follow-up plasma tHcy and provided adenoma data; 392 had one or more recurrent adenomas and 479 had no recurrent adenomas. The baseline characteristics of the study population by adenoma recurrence status are shown in Table 1. Those with an adenoma recurrence were significantly older; they were more likely to be male, have a higher BMI, and drink alcohol, and had lower plasma vitamin B₆ levels. Baseline plasma total folate was inversely associated with recurrence risk in the total study population (P = 0.013). Baseline plasma tHcy was significantly higher in those with an adenoma recurrence in the crude analysis (P = 0.04).

Baseline plasma tHcy decreased significantly over the 5-year recruitment period (*P* for trend < 0.001; data not shown). Mean baseline plasma tHcy was $10.7 \pm 3.6 \,\mu$ mol/L among subjects recruited in 1995 and decreased to a mean of $8.7 \pm 2.0 \,\mu$ mol/L among subjects recruited in 1998. By the time of the year 3 measurement mean plasma tHcy for those in the placebo group had leveled off at approximately 9.2 μ mol/L (*P* for trend 0.142).

Table 2 shows the univariate associations between selected baseline factors and plasma tHcy. Those in the highest quartile of tHcy were older and were more likely to be male, current smokers, and alcohol drinkers. They also had a higher BMI, lower plasma folate, B_2 , B_{12} , and B_6 , and a higher number of adenomas at the baseline examination. Table 3 shows the association between baseline plasma tHcy and the number of adenomas at the baseline examination. There was a borderline significant direct association between increasing plasma tHcy and the number of adenomas at the baseline examination after adjustment for age, sex, recruitment center, current smoking, BMI, and alcohol intake (P for trend = 0.06).

Subjects with the highest plasma tHcy were about 50% more likely to be in a higher baseline adenoma group. The proportional OR was 1.49 (95% CI, 0.95-2.31; *P* for trend = 0.06). After additional control for plasma folate

Table 1. Baseline characteristics of study subjects by adenoma recurrence						
Characteristic	Adenoma recurrence	No adenoma recurrence	P			
No. participants	392 (45.0)	479 (55.0)				
Mean age at baseline (SD), y	59.1 (9.4)	56.2 (9.5)	<0.001*			
Male sex, n (%)	270 (68.9)	289 (60.3)	0.01 [†]			
Colorectal cancer in first-degree relative, n (%)	117 (37.6)	148 (38.1)	0.91 [†]			
NSAID use at baseline, n (%)	319 (81.4)	379 (79.1)	0.41 [†]			
Multivitamin use at baseline, n (%)	134 (34.2)	183 (38.2)	0.22 [†]			
Race/Ethnicity, n (%)						
White	346 (88.3)	408 (85.2)	0.18 [†]			
African American	23 (5.9)	25 (5.2)				
Hispanic	16 (4.1)	27 (5.6)				
Other	7 (1.8)	19 (4.0)				
BMI, kg/m ²						
<25	104 (26.6)	161 (33.7)	0.08 [†]			
25-<30	191 (48.9)	212 (44.4)				
≥30	96 (24.6)	105 (22.0)				
Current cigarette smoker, n (%)	72 (18.5)	51 (10.7)	0.00 [†]			
Mean dietary intake (SD), kcal/day	1,645 (697)	1,614 (612)	0.49*			
Mean dietary folate intake (SD),* µg/day	316 (144)	327 (158)	0.31*			
Mean total folate intake (SD), µg/day	452 (256)	466 (249)	0.41*			
Mean baseline plasma folate (SD), nmol/L	22.1 (15.3)	25.3 (19.4)	0.01 [‡]			
Mean baseline plasma B ₁₂ (SD), pmol/L	322 (144)	337 (179)	0.15 [‡]			
Mean baseline plasma B ₆ (SD), nmol/L	77.8 (90.8)	84.1 (90.6)	0.01 [‡]			
Mean plasma Hcy at baseline (SD), µmol/L	10.1 (3.1)	9.6 (2.8)	0.04 [‡]			
Alcohol use at baseline, n (%)	278 (72.8)	297 (64.7)	0.01 [†]			
Caffeine use at baseline, n (%)	366 (95.8)	428 (93.3)	0.11 [†]			
History of high cholesterol at baseline, $n (\%)^{\$}$	142 (36.3)	147 (30.8)	0.08 [†]			
History of hypertension at baseline, $n \ (\%)^{\$}$	97 (24.7)	120 (25.1)	0.92 [†]			
Lifetime adenomas at baseline, mean (SD)	2.8 (2.6)	2.0 (1.7)	<0.001‡			
Number of baseline adenomas, mean (SD)	1.7 (1.1)	1.5 (0.9)	<0.001‡			
Advanced adenomas at baseline, $n (\%)^{\parallel}$						
0	248 (70.5)	315 (70.3)	0.97†			
≥1	104 (29.6)	133 (29.7)				
MTHFR 677 C>T genotype						
CC	178 (47.2)	197 (44.0)	0.35 [†]			
CT	166 (44.0)	199 (44.4)				
Π	33 (8.8)	52 (11.6)				
MTHFR 1298 A>C genotype						
AA	174 (46.0)	234 (52.1)	0.22 [†]			
AC	169 (44.7)	179 (39.9)				
CC	35 (9.3)	36 (8.0)				

Abbreviation: NSAID, nonsteroidal anti-inflammatory drugs.

*Two sample *t*-test.

 $^{\dagger}\chi^{2}$ test.

[‡]Nonparametric Wilcoxon rank sum test.

[§]Self-reported from the risk factor questionnaire.

^{||}Advanced adenoma was defined as a \geq 25% villous component, large adenoma (\geq 1 cm), advanced dysplasia, carcinoma *in situ*, or invasive cancer.

Characteristic*	Baseline plasma homocysteine, µmol/L [†]						
	≤7.95	7.96-9.26	9.27-11.25	>11.25	P [‡]		
Mean age (SD)	54.6 (9.3)	58.1 (9.1)	58.4 (9.6)	58.9 (9.7)	<0.001		
Male (%)	41.0	62.2	76.9	77.7	<0.001		
Current smoker (% yes)	12.2	11.7	11.9	21.0	0.01		
BMI (kg/m ²) %/category							
1 (≤25 kg/m²)	37.8	34.7	25.1	23.8	0.01		
2 (25-< 30 kg/m ²)	38.7	43.2	49.3	54.7			
3 (≥30 kg/m²)	23.4	22.1	25.6	21.5			
Alcohol drinker (% yes)	62.0	70.1	67.7	73.9	0.06		
Alcohol mean (SD), drinks/day among drinkers	0.6 (0.8)	0.9 (1.3)	0.9 (0.9)	1.2 (1.4)	<0.001		
Mean baseline plasma folate (SD), nmol/L	31.0 (18.5)	26.2 (16.6)	20.7 (14.0)	17.3 (17.6)	<0.001		
Mean baseline plasma B ₁₂ (SD), pmol/L	371 (145)	346 (210)	309 (113)	295 (163)	<0.001		
Mean baseline plasma B ₆ (SD), nmol/L	107 (118)	84 (87)	73 (84)	61 (57)	<0.001		
Mean baseline plasma B ₂ (SD), nmol/L	36 (50)	33 (61)	27 (51)	24 (43)	<0.001		
Number of adenomas at baseline exam, mean (SD)	1.4 (0.8)	1.5 (0.8)	1.7 (1.2)	1.7 (1.0)	0.001		
MTHFR 677 C>T genotype (% yes)							
CC	49.5	45.2	49.0	37.9	0.11		
CT	44.1	42.9	40.4	50.0			
TT	6.4	12.0	10.6	12.1			

Table 2. Univariate associations between selected risk factors and baseline plasma Hcy

*All risk factor values were taken from the baseline questionnaire.

[†]Quartile cut points based on the distribution of plasma tHcy in the whole study population.

[‡]*P* for trend for continuous variables (age, number of alcohol drinks, plasma levels, number of baseline adenomas) or *P* for heterogeneity for categorical variables (sex, current smoker, BMI, alcohol drinker, *MTHFR 677*).

and vitamins B_2 , B_6 , and B_{12} the adjusted proportional OR was attenuated at 1.18 (95% CI, 0.74-1.86; *P* for trend = 0.43).

Table 4 shows the association between baseline plasma tHcy and subsequent adenoma risk after stratifying on folic acid treatment group. The quartile 4/quartile 1 adjusted RRs were 0.98 (95% CI, 0.70-1.38; *P* for trend = 0.17) and 0.81 (95% CI, 0.58-1.12; *P* for trend = 0.17) in the placebo and supplemented groups, respectively (*P* for heterogeneity between treatment groups = 0.53). There was also no material difference in the RRs for advanced lesions (Table 4). There was no modification of RRs for either end point by sex, aspirin treatment group, or *MTHFR 677C>T* genotype (data not shown).

Discussion

In this analysis of subjects participating in a randomized clinical trial of folate and/or aspirin for the prevention of colorectal adenomas there was no association between baseline plasma tHcy and adenoma recurrence risk in either the placebo or the folic acid supplementation groups. The lack of association between plasma tHcy and recurrence risk was similar for all adenoma end points. Baseline plasma tHcy was associated with the number of adenomas at the baseline examination, but this association was attenuated and no longer statistically significant after controlling for potential confounders, including plasma total folate and other B vitamins.

Table 3. Association between baseline plasmatHcy and number of adenomas at the baselineexam

Baseline homocysteine (µmol/L)	Adjustor set A* OR (95% CI) [‡]	Adjustor set B [†] OR (95% CI)		
≤7.95	Reference	Reference		
7.96-9.26	1.15 (0.74-1.79)	1.02 (0.63-1.66)		
9.27-11.25	1.37 (0.87-2.16)	1.17 (0.73-1.88)		
>11.25	1.49 (0.95-2.31)	1.18 (0.74-1.86)		
	<i>P</i> trend = 0.06	<i>P</i> trend = 0.43		

*Adjustor set A consists of age, sex, study center, alcohol (continuous), BMI (continuous), and current smoking.
[†]Adjustor set B consists of age, sex, study center, alcohol (continuous), BMI (continuous), current smoking, plasma B₂, plasma B₆, plasma B₁₂, and plasma folate.
[‡]OR calculated using a proportional odds model where the outcome is number of baseline adenomas (1, 2, 3, 4, or 5+).

Plasma tHcy µmol/L	Placebo group			P_{trend}		Folate group				
	≤7.95	7.96-9.26	9.27-11.25	>11.25		≤7.95	7.96-9.26	9.27-11.25	>11.25	
All adenomas										
Recurrences/total	41/109	57/118	40/102	49/102		44/113	53/104	55/110	53/113	
Adjusted RR	1.0	1.15	0.84	0.98	0.17	1.0	1.05	0.95	0.81	0.17
(95% CI)*		(0.84-1.58)	(0.60-1.19)	(0.70-1.38)			(0.76-1.44)	(0.69-1.32)	(0.58-1.12)	
P heterogeneity					0.53					
Advanced adenomas [†]										
Recurrences/total	7/103	13/115	10/101	8/101		13/111	13/102	7/110	19/110	
Adjusted RR	1.0	1.39	1.20	1.00	0.83	1.0	0.86	0.41	0.98	0.21
(95% CI)*		(0.56-3.42)	(0.46-3.13)	(0.36-2.75)			(0.40-1.86)	(0.17-1.03)	(0.47-2.07)	
P heterogeneity					0.30					

Table 4. Association between baseline plasma tHcy and adenoma recurrence risk by folate treatment group and adenoma type

*Adjusted for age, sex, study center, alcohol (continuous), BMI (continuous), smoking status (never, former, current), plasma B₂, plasma B₆, plasma B₁₂, plasma folate, and aspirin treatment assignment.

[†]Advanced adenoma was defined as \geq 25% villous adenoma, large adenoma (\geq 1 cm), advanced dysplasia, carcinoma *in situ*, or invasive cancer.

Homocysteine is formed from SAH, a product of transmethylation reactions using *S*-adenosylmethionine (SAM) as methyl donor. Hcy can be remethylated to methionine or, alternatively, irreversibly transsulfurated to cystathionine by the vitamin B_6 -dependent enzyme cystathionine- β -synthase. Any Hcy that is not remethylated or converted to cystathionine is rapidly exported to plasma where it can be measured as a combination of related protein-bound thiols and free Hcy, referred to as plasma total Hcy (plasma tHcy; ref. 31).

Plasma tHcy has been suggested to be a direct measure of intracellular methylation capacity (11, 13, 32) due to its equilibrium with SAH, a potent inhibitor of SAM-dependent methylation reactions. In the study by Yi et al. (13) the relationship between plasma tHcy and SAH seemed to be linear for plasma tHcy values from 5 to 18 µmol/L, suggesting a positive association even for very low values of plasma tHcy. That this association may be functionally relevant is suggested by several studies that have reported significantly less global DNA methylation in circulating lymphocytes or colorectal epithelium in those with higher plasma tHcy (13, 17, 33), and in human umbilical vein endothelial cells (32). However, none of the human studies have shown this association to be independent of plasma folate levels. Plasma tHcy may also be associated with increased DNA damage. Fenech et al. (15) reported that plasma tHcy >10 µmol/L was associated with increased numbers of micronuclei in circulating lymphocytes, an association independent of circulating folate. Additionally, plasma tHcy is a potentially more inclusive marker of compromised one-carbon metabolism than is plasma folate alone because of its association with B vitamins other than folate (1-4), as well as the MTHFR 677C>T genotype (5-8). In one study plasma

tHcy was a more sensitive index of colonic mucosal folate than was plasma folate (34).

Previous studies of the association between plasma tHcy and risk of colorectal neoplasia have had mixed results. Four studies reported increased risk of colorectal cancer (12, 16-18), colorectal adenomas (17), or adenoma recurrence (12) for those with higher plasma tHcy. However, the increase was statistically significant in only two of these studies (12, 18) and only one (12) controlled for plasma folate. Our results are concordant with those of three other recent studies that did not observe a significant increase in colorectal cancer risk with increasing plasma tHcy (19-21).

We did not observe any association between plasma tHcy and adenoma recurrence even among subjects assigned to placebo. About half the subjects in our study were recruited after voluntary folate fortification of the U.S. food supply began in 1996, and the first 3-year observation period overlapped a time of gradually increasing folic acid availability in U.S. and Canadian diets, with consequently decreasing tHcy levels. It is possible that our negative results are due to the progressively lower plasma tHcy, which may have fallen to levels below a threshold for an association with adenoma risk. In this regard, it is of interest that one of the studies that did not find an association of tHcy with colorectal cancer risk (19) relied on blood samples collected from 2001 to 2006, after fortification of grain products with folic acid, as in the current study. However, this would not explain the null results of two other studies (20, 21) that involved nonfortified European populations with comparatively higher plasma tHcy levels.

Plasma tHcy levels were highest at the baseline examination. The number of adenomas at the baseline

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examination was significantly associated with baseline plasma tHcy in univariate analysis. When we pursued this finding further, we found that after controlling for other plasma tHcy determinants (e.g., plasma folate and other B vitamins) the association between baseline plasma tHcy and adenomas was substantially attenuated and no longer statistically significant. This suggests that even at the higher plasma tHcy levels at the baseline examination there was little independent effect of plasma tHcy in this study population.

This study has several limitations, including the requirement that all subjects in the study cohort have at least one adenoma before randomization, a design feature that limits the generalizability of our results, and the progressively higher folate levels our study subjects experienced over the period of follow-up, even for those randomized to placebo. We were not able to control for serum creatinine, a well-known determinant of plasma tHcy. However, we do not think this affected the main result of no association between plasma tHcy and adenoma recurrence because creatinine is not a risk factor for either adenoma occurrence or recurrence and therefore is not likely to be a confounder by definition. For the analysis of the association of baseline plasma tHcy and the number of baseline adenomas we would not expect confounding *a priori* but in the worst case scenario, where increased serum creatinine is positively associated with baseline adenoma number as well as higher plasma tHcy, the bias would be away from the null toward an increased risk for multiple adenomas in those with higher plasma tHcy as we saw in the minimally adjusted model. However, our results were null in the fully adjusted model, suggesting that this was not a major source of confounding in the final analysis. Additionally, because baseline plasma tHcy was measured after the occurrence of prior adenomas we cannot exclude the possibility that the presence of adenomas affected baseline plasma tHcy, although this seems unlikely.

The advantages of our study include the large sample size and the high follow-up rate for subjects. Additionally,

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we had data on many known determinants of plasma tHcy and could control for them in our analysis. Finally, because of the prospective design and the fact that all subjects were cleared of polyps at their baseline exam, we were able to assess the effect of baseline plasma tHcy on incident as well as prevalent adenomas.

In summary, we did not see an association between higher baseline plasma tHcy and adenoma recurrence risk in this prospective study of folate and aspirin supplementation. These results do not support those of some earlier studies, which suggested that plasma tHcy may be a marker for increased risk of colorectal cancer, adenoma, and adenoma recurrence at higher plasma tHcy values, but are concordant with three other studies that did not observe any association between plasma tHcy and colorectal cancer risk. Our data suggest one of two possibilities: that there is no independent association between plasma tHcy and adenoma recurrence risk or that any association between plasma tHcy and adenoma recurrence may be limited to plasma tHcy levels higher than those characterizing the current, largely folic acidfortified, study population.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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